

# MaxFection<sup>™</sup> 8600 in vitro Transfection Reagent

MF8600-015 for  $\sim$ 200 in vitro transfections Store at -20 °C

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# Description

MaxFection<sup>™</sup> 8600 belongs to an efficient new class of non-viral gene delivery based on biodegradable cationic polymer.

# **Principle**

The remarkable transfection efficiency of MaxFection<sup>™</sup> 8600 is due to its intracellular degradation property, which may maximize gene unloading into nucleus as well as minimize cytotoxicity of cationic residues.

# **Reagent Supplied**

- 1) 10×MaxFection<sup>™</sup> 8600 concentrated solution (0.45mL, =4.5mL 1X reagent)
- 2) Buffer solution (>2mL)

#### **Formulation**

MaxFection<sup>™</sup> 8600 is sterile apyrogenic solution of cationic polymer in an acidic pH 5.0 buffer.

Considerations for Transfection with MaxFection™ 8600

#### 1. DNA quality

High quality DNA is very important for successful transfection.

DNA should be sterile and free of any contaminant such as endotoxins.

The OD<sub>260</sub>/OD<sub>280</sub> ratio should be at 1.6 or greater.

# 2. Cell density at transfection

The recommended cell density (confluency) for most cell types is 50-70% for transfection. The cells should not be confluent or at stationary phase prior to transfection.

#### 3. Transfection incubation time

Detection of transgene expression should be performed within 24-72 hours. An optimal post-transfection incubation time can be determined using a reporter gene (such as Luciferase, galactosidase or Green Fluorescent Protein).

### 4. Choice of promoter

High transfection efficiency depends not only on cell line type but promoter under which the gene of interest is expressed. MaxFection™ 8600 may be applied for cytomegalovirus (CMV) promoter, one of the best known for high transgene expression in a wide variety of cell lines, and others such as ORF promoter.

# 5. MaxFection<sup>™</sup> 8600/DNA equivalent

- The required amount of MaxFection<sup>™</sup> 8600 solution depends on the amount of DNA and the number of equivalents needed.
- Initially, we recommend to use 1  $\mu$ g of DNA and 15  $\mu$ l of 1× MaxFection<sup>TM</sup> reagent per well of 24-well plates (see Table 1).
- Subsequent optimization may further increase the transfection efficiency in your transfection test depending on cell line type and the gene plasmid.



• The DNA quantity can range from 0.5 to 10 µg for 100,000 cells; likewise MaxFection™ 8600/DNA ratio can range from 15 to 40uL 1X .

### 6. Transfection in the presence of serum

MaxFection <sup>™</sup> 8600-mediated high transfection efficiency is not seriously affected by 10% serum.

# 7. Protocol for transfection of adherent cells in a 24-well plate.

Quantities and volumes should be scaled up according to the number of cells or wells to be transfected. (See Table 1 for scale-up ratios).

- 1) Prepare DNA working solution: Dilute 1µg of DNA with *Buffer solution* in a 1.5 mL of EP tube to give 10 µL of DNA working solution;
- 2) Prepare  $1 \times MaxFection^{TM}$  8600 solution: Dilute 3  $\mu L$  of  $10 \times MaxFection^{TM}$  8600 concentrated solution with 27  $\mu L$  of *Buffer solution*; You may scale up as needed.
- 3) Add 30  $\mu$ L of 1×MaxFection<sup>TM</sup> 8600 solution into 20  $\mu$ L of DNA working solution and vortex-mix (or pipette) the solution **immediately** for 5 sec;
- 4) Incubate for 30 minutes at room temperature;
- 5) Add 25 µl of the MaxFection™ 8600/DNA mixture to each well;
- 6) Gently rock the plate back and forth and from side to side to achieve even distribution of the complexes;
- 7) Incubate at 37°C for 1-4 hour transfection.
- 8) For serum-free transfection, replace serum-free culture medium with complete culture medium
- 9) Monitor transfection after 24-72 h post-transfection **Notes.** 1. Initially, we recommend the use of 1 μg DNA and 15 μL 1×MaxFection<sup>TM</sup> 8600 per well of 24-well plate (see Table 1) in **serum-free transfection for 1-4 h and 24-72 h post-transfection in complete**

#### medium.

2. Subsequent optimization may increase the equivalent number from 20-100 depending on the cell line and the gene expressed.

**Table 1.** Scale-up equivalent numbers were used according to the surface area of the tissue culture plate

Culture Plate	Growth Area (cm²/well)	Cell seeding density	Recommended DNA amount (µg)	optimal amount of DNA (µg)*	µL of 1×MaxFection™ 8600 (µL)
96	0.3	0.5-1×10 <sup>4</sup>	0.3-1	0.5	7.5
48	0.7	1-3×10 <sup>4</sup>	0.5-1	0.5	7.5
24	2	2-6×10 <sup>4</sup>	1-1.5	1	15
12	4	0.4-1.2×10 <sup>5</sup>	1-3	1.5	22.5
6	9.5	1-2×10 <sup>5</sup>	2-4	2	30

Actual values depend on the cell line type

# Cell lines successfully transfected with MaxFection™ 8600 include:

Permanently growing cell lines				
HeLa-	Human	Cervix epitheloid carcinoma cells		
MCF-7	Human	Breast adenocarcinoma cells		
PC-3	Human	Prostate adenocarcinoma cells		
SKOV-3	Human	Ovarian carcinomacells		
HepG2	Human	Hepatoma cells		
A549	Human	Type II pneumocytes		
NIH 3T3	Mouse	Embrionic fibroblasts		
COS-7	Monkey	African green monkey kidney cells		